

RESEARCH PROPOSAL



UNIVERSITAS
INDONESIA

Veritas, Probitas, Iustitia
— EST. 1849 —

**THE ROLE OF GUT MICROBIOME AND CHRONIC INFLAMMATION IN
YOUNG-ONSET COLORECTAL CANCER**
Next-Generation Sequencing (NGS) as screening method

RESEARCH TEAM
MURDANI ABDULLAH
SAFARINA G MALIK
ARI FAHRIAL SYAM
YUSRA
FAUZI YUSUF

FACULTY OF MEDICINE
UNIVERSITAS INDONESIA

24 April 2019

INTRODUCTION

Background

In 2025 the World Health Organization (WHO) estimates that cancer will have a higher mortality rate than coronary heart disease and stroke. It is estimated there are more than 20 million new cases of cancer each year in low- and middle-income countries due to global epidemiological changes. Colorectal cancer is the third most prevalent cancer worldwide and is in the fourth place in cancer mortality rate. The incidence of colorectal cancer is estimated to be 1.2 million each year with more than 630.000 number of deaths, accounting for 8% of deaths caused by cancer. In 2030 it is estimated there will be 2.2 million number of new cases and 1.1 million number of deaths caused by colorectal cancer. Based on Jakarta Cancer Registry in 2012 in Indonesia, colorectal cancer is the second most common cancer in men and the fourth most common in women. The incidence of colorectal cancer was 3.15 per 100.000 of populations in women and 4.13 per 100.000 of populations in men.

Previous studies indicated that colorectal cancer patients in Indonesia were younger compared to developed countries and were sporadic. Sporadic colorectal cancer generally resulted from somatic mutation without any correlation to family history. In comparison to patients in developed countries, colorectal cancer patients in Indonesia also showed differences in several characteristics including distal localization (rectum), higher clinical staging, and poor prognosis.

Increasing number of colorectal cancer cases make a profound impact on national health expenditures. In the era of *Jaminan Kesehatan Nasional / JKN* (the National Health Insurance in Indonesia), there are limitations on budgets for colorectal cancer management. Thus, numerous efforts are needed on the early screening and prevention of colorectal cancer.

Gastrointestinal system comprises of more than 200 m² of mucosal surface. The immune system in gastrointestinal mucosa has been studied extensively until this day. The microflora or the microorganism in gastrointestinal system, also commonly known as the gut microbiome, is a part of the immunity system in gastrointestinal mucosa. Around 20% of malignancy are linked to microorganism. The gut microbiome is suspected to have a role in genetic and epigenetic changes that cause dysplasia, clonal expansion, tumor growth, and cancer. In addition, chronic

inflammation is also suspected to have a role in colorectal carcinogenesis. One of the chronic inflammation markers linked with carcinogenesis is NF- κ B.

Several studies showed that dysbiosis or imbalance of gut microbiome is linked with mucosal barrier damage, chronic inflammation and production of carcinogenic metabolite causing neoplasm. Thus far, the association between gut microbiome and colorectal cancer has provided an opportunity to develop new strategies in the screening method of colorectal cancer.

Non-invasive methods for colorectal cancer screening are currently being developed to replace colonoscopy. One of the screening methods is *fecal immunochemical test* (FIT) using fecal samples. This method has a sensitivity of 87.5% in detecting colorectal cancer cases. Another screening method uses blood sample to evaluate *carcinoembryonic antigen* (CEA) level in blood serum. An increase in CEA serum level of more than 5 ng/mL is found in 17-47% of colorectal patients and is associated with poor prognosis.

Research Objectives

General Objectives

To investigate the role of gut microbiome pattern and inflammation marker NF- κ B in colorectal cancer patients

Specific Objectives

1. To obtain data on gut microbiome pattern on young-onset colorectal cancer patients and colorectal cancer patients aged >50 years old.
2. To explore the role of inflammation marker NF- κ B on young-onset colorectal cancer patients and colorectal cancer patients aged >50 years old.
3. To obtain CEA values of young-onset colorectal cancer patients and colorectal cancer patients aged >50 years old.
4. To obtain data on gut microbiome which could be used as screening methods in patients suspected with colorectal cancer

Hypothesis

1. There are differences in gut microbiome pattern on young-onset colorectal cancer patients and colorectal cancer patients aged >50 years old compared to healthy individuals
2. There are differences in NF- κ B on young-onset colorectal cancer patients and colorectal cancer patients aged >50 years old compared to healthy individuals
3. There are differences in CEA values on young-onset colorectal cancer patients and colorectal cancer patients aged >50 years old compared to healthy individuals

Research Benefits

For Researchers

Researchers will expand their knowledge on gut microbiome pattern in colorectal cancer patients.

For Institutions

This study will provide a contribution of science in national and international scale. The result of this study could be used as a guideline on utilizing gut microbiome data on colorectal cancer management.

For Communities

The result of this study is expected to improve the management of colorectal cancer as a screening method that would be applicable in daily clinical practice.

METHODOLOGY

Study Design

This is a cross-sectional study.

Time and Place

This study is conducted in Dr. Cipto Mangunkusumo Hospital: Digestive Endoscopy Center, Department of Internal Medicine, Department of Clinical Pathology, Department of Anatomical Pathology, and Eijkman Institute of Molecular Biology from May 2019 to May 2022.

Population

Target Population

Patient suspected with colorectal cancer in Indonesia.

Accessible Population

Patients suspected with colorectal cancer who come to Digestive Endoscopy Center in Dr. Cipto Mangunkusumo Hospital to conduct colonoscopy procedure and histopathological examination.

Subject Criteria

Inclusion Criteria

1. Age \geq 35 years old
2. Suspected with colorectal cancer and undergoing a colonoscopy procedure
3. No history of colorectal cancer treatment

Exclusion Criteria

1. Unwilling to provide fecal and blood sample
2. Incomplete colonoscopy procedure due to any reasons

Estimated Sample Size

Since this is a pilot study to obtain gut microbiome pattern on colorectal cancer patients, the number of samples is determined by the researchers: 100 subjects for neoplasm and 50 subjects for non-neoplasm (according to histopathology report).

Randomisation

The sampling will not be randomized. All patients suspected with colorectal cancer who come to Digestive Endoscopy Center in Dr. Cipto Mangunkusumo Hospital to conduct colonoscopy procedure and histopathological examination who fulfilled the criteria will be recruited for this study and will undergo a series of examinations

Operational Definition

No.	Variable	Definition	Source	Scale
1.	Patient suspected with colorectal cancer	Patient suspected with colorectal cancer by clinician based on clinical findings such as gastrointestinal bleeding, chronic constipation, chronic diarrhea, nonspecific abdominal pain, weight loss, palpable abdominal mass	Clinical judgement	Nominal
2.	Age	Patient's age (in years)	Medical record	
3.	Sex	Patient's sex	Medical record	Nominal
4.	Family history of colorectal cancer in a first-degree relative	First-degree relative: parents, sibling, children	Medical record and history taking	Nominal 1= No 2= Yes
5.	History of smoking	Past or current history of smoking	Medical record and history taking	Nominal 1= No 2= Yes
4.	FIT results	The result of Fecal immunochemical test (FIT)	Fecal immunochemical test (FIT)	Nominal 1= Negative 2= Positive
5.	Gut microbiome results	The result of gut microbiome examination	Next Generation Sequencing (NGS)	Nominal 1: Fusobacterium 2: Bacteroides 3. Lachnospiraceae

6.	NF- κ B results	The results of NF- κ B examination: positive if accumulated score ≥ 3	Examination with immune-histochemical method	Nominal 1= Negative 2= Positive
7.	CEA results	The results of Carcinoembryonic antigen (CEA) examination	Examination with ELISA method	Nominal 1= Negative 2= Positive
7	Histo-pathological results	The results of histopathological examination from biopsy samples are used as gold standard of colorectal cancer diagnosis	Medical record	Nominal 1 = Colorectal cancer 2 = Non-colorectal cancer

Ethical Approval

This study will be obtained ethical approval from the Ethics Committee of Faculty of Medicine, University of Indonesia.

Research Flow

Patients suspected with colorectal cancer who come to Digestive Endoscopy Center in Dr. Cipto Mangunkusumo Hospital to conduct colonoscopy procedure and histopathological examination who fulfilled the criteria will be recruited for this study and will undergo a series of examinations. Each patient will be given an explanation of this study and will be asked his/her willingness to participate on this study by signing the informed consent form. Baseline characteristics data will be obtained from medical record. Before the colonoscopy procedure is conducted, each patient will undergo a score assessment, blood sampling, and fecal sampling.

1. Score Assessment

Asia Pacific Colorectal Screening (APCS) is a validated tool to predict the risk of colorectal cancer in asymptomatic Asian population. The scoring system comprises of three categories: low risk (score 0-1), moderate risk (score 2-3) and high risk (score 4-7). Patients with moderate and high risk will undergo further examinations

Table. Asia Pacific Colorectal Screening (APCS).

Risk Factor	Criteria	Score
Age (in years)	< 50	0
	50 – 69	2
	≥ 70	3
Sex	Female	0
	Male	1
Family history of colorectal cancer in a first-degree relative	Absent	0
	Present	2
History of smoking	Never	0
	Current or past	1

2. Blood Sampling

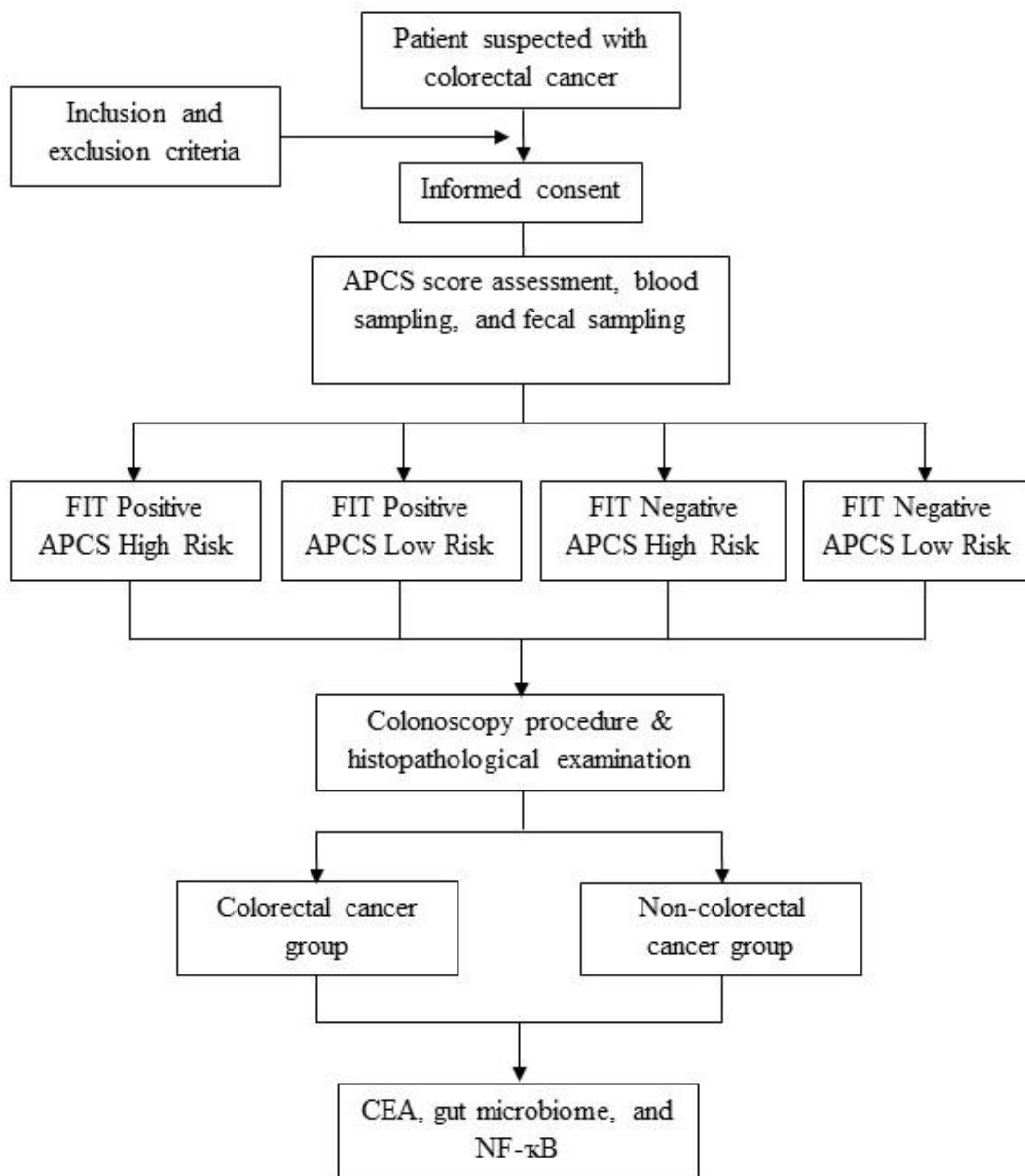
Blood samples will be taken before colonoscopy procedure to evaluate the level of serum CEA by ELISA method and to evaluate the presence of NF- κ B by immunohistochemical method.

- a. Carcinoembryonic antigen (CEA) is a well-known marker for colorectal cancer. A pre-treatment serum CEA level of ≥ 5 ng/mL is associated with poor prognosis in colorectal cancer patients.
- b. NF- κ B is a chronic inflammation marker found in colorectal cancer patients

3. Fecal Sampling

Fecal samples will be taken before colonoscopy procedure to be tested for FIT and to evaluate the gut microbiome.

- a. Fecal immunochemical test (FIT) is a recommended screening method for colorectal cancer. Detection of hemoglobin over a certain level in fecal samples indicated a positive FIT. Patients with positive FIT will undergo further examinations.
- b. Gut microbiome examination will be conducted with next generation sequencing (NGS) method.



Blood samples and fecal samples will be tested in Department of Clinical Pathology Dr. Cipto Mangunkusumo Hospital, while gut microbiome will be evaluated in Eijkman Institute of Molecular Biology. Colonoscopy procedure will be conducted in Digestive Endoscopy Center Dr. Cipto Mangunkusumo Hospital. Histopathological examination be conducted in Department of Anatomical Pathology Dr. Cipto Mangunkusumo Hospital. The results of colonoscopy procedures and histopathology report will be obtained from medical record.

Gut microbiome examination from fecal samples will be initiated by DNA isolation. After extraction material is obtained, PCR will be performed with 16S RNA Sequencing. Metagenomic study will be carried out by analysing prokaryotic 16S ribosomal RNA gene (16S rRNA) which has a length of 1500 base pairs and consists of 9 variable regions that are between conserved regions. Variable regions that are generally used for phylogenetic analysis are V3 and V4 for classification to the level of genus or species. To obtain gut microbiome pattern, Next-Generation Sequencing (NGS) examination will be conducted.

TIMELINE

First Year

No	Activities	Month											
		1	2	3	4	5	6	7	8	9	10	11	12
1	Obtaining ethical approval	X	X										
2	Obtaining location permit		X	X									
3	Sampling				X	X	X	X	X	X	X	X	X
4	FIT, CEA, NF-kB, gut microbiome examinations				X	X	X	X	X	X	X	X	X

Second Year

No	Activities	Month											
		1	2	3	4	5	6	7	8	9	10	11	12
1	Sampling	X	X	X									
2	FIT, CEA, NF-kB, gut microbiome examinations	X	X	X									
3	Data analysis and report writing				X	X	X	X	X	X	X	X	X

Third Year

No	Activities	Month											
		1	2	3	4	5	6	7	8	9	10	11	12
1	Data analysis and report writing	X	X	X									
2	Scientific publication		X	X	X	X	X	X	X	X	X	X	X

REFERENCES

1. Ferlay J, Soerjomataram I, Dikshit R, et al. Cancer incidence and mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*. 2015;136(5):E359-E386. doi:10.1002/ijc.29210
2. Arnold M, Sierra MS, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global patterns and trends in colorectal cancer incidence and mortality. *Gut*. 2017;66(4):683-691. doi:10.1136/gutjnl-2015-310912
3. Moghimi-Dehkordi B. An overview of colorectal cancer survival rates and prognosis in Asia. *World J Gastrointest Oncol*. 2012;4(4):71. doi:10.4251/wjgo.v4.i4.71
4. Indonesia KKKK. Paduan Penatalaksanaan Kanker Kolorektal. In: *Kanker Kolorektal*. ; 2014:xi+146 hlm.
5. Wahidin M, Noviani R, Hermawan S, Andriani V, Ardian A, Djarir H. Population-Based Cancer Registration in Indonesia. *Asian Pacific J Cancer Prev*. 2012;13(4):1709-1710.
6. Abbas AK, Lichtman AH, Pillai S. *Specialized Immunity at Epithelial Barriers and in Immune Privileged Tissues*. 9th ed. Philadelphia: Elsevier Inc.; 2018.
7. Oke S, Martin A. Insights into the role of the intestinal microbiota in colon cancer. *Ther Adv Gastroenterol Rev*. 2017;10(5):417-428. doi:10.1177/ https://doi.org/10.1177/17562
8. Raskov H, Burcharth J, Pommergaard HC. Linking gut microbiota to colorectal cancer. *J Cancer*. 2017;8(17):3378-3395. doi:10.7150/jca.20497
9. Chen H, Yu Y, Wang J, et al. Decreased dietary fiber intake and structural alteration of gut microbiota in patients with advanced colorectal adenoma. *Am J Clin Nutr*. 2013;97:1044-1052.
10. Louis P, Hold GL, Flint HJ. The gut microbiota, bacterial metabolites and colorectal cancer. *Nat Rev Microbiol*. 2014;12(10):661-672.
11. Ai L, Tian H, Chen Z, Chen H, Xu J, Fang J-Y. Systematic evaluation of supervised classifiers for fecal microbiota-based prediction of colorectal cancer. *Oncotarget*. 2017;8(6):9546-9556. doi:10.18632/oncotarget.14488
12. Tarantino I, Warschkow R, Schmied BM, et al. Predictive Value of CEA for Survival in Stage I Rectal Cancer : a Population-Based Propensity Score-Matched Analysis. *J*

- Gastrointest Surg.* 2016;1213-1222. doi:10.1007/s11605-016-3137-8
13. Probst CP, Becerra AZ, Aquina CT, et al. Watch and Wait ? — Elevated Pretreatment CEA Is Associated with Decreased Pathological Complete Response in Rectal Cancer. *J Gastrointest Surg.* 2015. doi:10.1007/s11605-015-2987-9
 14. Yoon SM, Kim DY, Kim TH, et al. Clinical Parameters Predicting Pathologic Tumor Response after Preoperative Chemoradiotherapy for Rectal Cancer. *Int J Radiat Oncol Biol Phys.* 2007;69(4):1167-1172. doi:10.1016/j.ijrobp.2007.04.047
 15. Polat E, Duman U, Duman M, et al. Diagnostic value of preoperative serum carcinoembryonic antigen and carbohydrate antigen 19-9 in colorectal cancer. 2014;21:1-7.
 16. Gao Y, Wang J, Zhou Y, Sheng S, Qian SY, Huo X. Evaluation of Serum CEA , CA19-9 , CA72-4 , CA125 and Ferritin as Diagnostic Markers and Factors of Clinical Parameters for Colorectal Cancer. *Sci Rep.* 2018;(October 2017):1-9. doi:10.1038/s41598-018-21048-y
 17. Trihartini P. Cancer Registration in Indonesia. 2001;2:21-24.
 18. Valentini V, Aristei C, Glimelius B, et al. Multidisciplinary Rectal Cancer Management: 2nd European Rectal Cancer Consensus Conference (EURECA-CC2). *Radiother Oncol.* 2009;92(2):148-163. doi:10.1016/j.radonc.2009.06.027
 19. Boyle P, Langman JS. ABC of colorectal cancer: Epidemiology. *BMJ Br Med J.* 2000;321(7264):805. doi:10.1002/bjs.1800581022
 20. Conteduca V, Sansonno D, Russi S, Dammacco F. Precancerous colorectal lesions (Review). *Int J Oncol.* 2013;43(4):973-984. doi:10.3892/ijo.2013.2041
 21. Abdullah M, Sudoyo AW, Utomo AR, Fauzi A, Rani AA. Molecular profile of colorectal cancer in Indonesia: Is there another pathway? *Gastroenterol Hepatol from Bed to Bench.* 2012;5(2):71-78.
 22. Effendi-ys R, Rey I. Molecular Diagnostic in Colorectal Cancer. *Indones J Gastroenterol , Hepatol Dig Endosc.* 2015;16:26-33.
 23. Luther J, Chan AT. Malignant tumour of the colon. In: Podolsky DK, Camilleri M, Fitz JG, Kalloo AN, Shanahan F, Wang TC, eds. *Yamada's Atlas of Gastroenterology.* 5th ed. West Sussex: Wiley & Sons Ltd.,; 2016:238-245.
 24. Sanford KW, McPherson RA. Fecal Occult Blood Testing. *Clin Lab Med.*

- 2009;29(3):523-541. doi:10.1016/j.cll.2009.06.008
25. Kim BSM. Diagnosis of gastrointestinal bleeding: A practical guide for clinicians. *World J Gastrointest Pathophysiol.* 2014;5(4):467. doi:10.4291/wjgp.v5.i4.467
 26. Wang T, Cai G, Qiu Y, et al. Structural segregation of gut microbiota between colorectal cancer patients and healthy volunteers. *ISME J.* 2012;6(2):320-329.
 27. Rowland IR. The role of the gastrointestinal microbiota in colorectal cancer. *Curr Pharm Des.* 2009;15(13):1524-1527. doi:10.2174/138161209788168191
 28. Rezasoltani S, Aghdaei HA, Mojarad EN, Dabiri H, Ghanbari R, Zali MR. Gut microbiota, epigenetic modification and colorectal cancer. *Iran J Microbiol.* 2017;9(2):55-63.
 29. Fujio-vejar S, Vasquez Y, Morales P, Magne F, Ward NL. The Gut Microbiota of Healthy Chilean Subjects Reveals a High Abundance of the Phylum Verrucomicrobia. 2017;8(June):1-11. doi:10.3389/fmicb.2017.01221
 30. Vaiopoulos AG, Athanasoula KC, Papavassiliou AG. NF- κ B in colorectal cancer. *J Mol Med.* 2013;91(9):1-9. doi:10.1007/s00109-013-1045-x
 31. Brennan CA, Garrett WS. Gut Microbiota, Inflammation, and Colorectal Cancer. *Annu Rev Microbiol.* 2016;70:395-411.
 32. González-Quezada BA, Francisco U, Bejarano S, et al. Expression profile of NF - κ B regulated genes in sporadic colorectal cancer patients. 2018:7344-7354. doi:10.3892/ol.2018.8201
 33. Lee JH, Lee SW. The Roles of Carcinoembryonic Antigen in Liver Metastasis and Therapeutic Approaches. *Gastroenterol Res Pract.* 2017;2017. doi:10.1155/2017/7521987
 34. Duffy MJ. Carcinoembryonic antigen as a marker for colorectal cancer: Is it clinically useful? *Clin Chem.* 2001;47(4):624-630. doi:10.1084/jem.121.3.439
 35. Jong Hoon L, Dae Yong K, Sung Hwan K, et al. Carcinoembryonic antigen has prognostic value for tumor downstaging and recurrence in rectal cancer after preoperative chemoradiotherapy and curative surgery : A multi-institutional and case-matched control study of KROG 14-12. *Radiother Oncol.* 2015;(116):202-208. doi:10.1016/j.radonc.2015.07.049
 36. Shinkins B, Nicholson BD, Primrose J, et al. The diagnostic accuracy of a single CEA blood test in detecting colorectal cancer recurrence: Results from the FACS trial. *PLoS*

One. 2017;12(3):1-11. doi:10.1371/journal.pone.0171810

37. Yeoh K-G, Ho K-Y, Chiu H-M, et al. The Asia-Pacific Colorectal Screening score: a validated tool that stratifies risk for colorectal advanced neoplasia in asymptomatic Asian subjects. *Gut*. 2011;60(9):1236-1241. doi:10.1136/gut.2010.221168