¹³C-butyrate and ¹³C-glucose breath testing to detect mesenteric ischemia, a proof of principal study in healthy volunteers (CMI breath test) (June 2020)

PROTOCOL TITLE ⁽¹³C-butyrate and ¹³C-glucose breath testing to detect mesenteric ischemia, a proof of principal study in healthy volunteers'

Protocol ID	Not applicable
Short title	CMI breath test
EudraCT number	Not applicable
Version	2.0
Date	08-06-2020
Coordinating investigator/project	L.G. Terlouw, MD
leader	Department of Gastroenterology & Hepatology,
	Erasmus MC University Medical Center, Rotterdam,
	the Netherlands
	<u>l.terlouw@erasmusmc.nl</u>
Principal investigator(s) (in	Erasmus MC University Medical Center
Dutch: hoofdonderzoeker/	M.J. Bruno, MD, PhD
uitvoerder) <multicenter per="" research:="" site=""></multicenter>	Department of Gastroenterology & Hepatology,
	Erasmus MC Universitv Medical Center. Rotterdam.
	the Netherlands
	m.bruno@erasmusmc.nl
	A. Moelker, MD, PhD
	Department of Radiology,
	Erasmus MC University Medical Center, Rotterdam,
	the Netherlands
	<u>a.moelker@erasmusmc.nl</u>
	M.P. Peppelenbosch, PhD
	Department of Gastroenterology & Hepatology,
	Erasmus MC University Medical Center, Rotterdam,

	the Netherlands
	m.peppelenbosch@erasmusmc.nl
	H.J.M. Verhagen, MD, PhD
	Department of vascular surgery,
	Erasmus MC University Medical Center, Rotterdam, the Netherlands
	h.verhagen@erasmusmc.nl
	T.M. Luider, PhD
	Department of Neurology,
	Erasmus MC University Medical Center, Rotterdam, the Netherlands
	<u>t.luider@erasmusmc.nl</u>
	Franciscus Gasthuis & Vlietland
	D. van Noord, MD, PhD
	Department of Gastroenterology & Hepatology,
	Franciscus Gasthuis & Vlietland, Rotterdam, the Netherlands
	<u>d.leemreis@franciscus.nl</u>
Sponsor (in Dutch:	Erasmus University Medical Center Rotterdam
verrichter/opdrachtgever)	
Subsidising party	
Independent expert (s)	A.C. de Vries, MD, PhD
	Department of Gastroenterology & Hepatology,
	Erasmus MC University Medical Center, Rotterdam,

	the Netherlands
	B.M. van Dalen, MD, PhD
	Department of Cardiology,
	Franciscus Gasthuis & Vlietland, Rotterdam,
	the Netherlands
	<u>b.vandalen@franciscus.nl</u>
Laboratory sites <if applicable=""></if>	Laboratorium MDL
	Erasmus MC University Medical Center,
	Wytemaweg 80,
	3015 CN Rotterdam, the Netherlands
	Da Vinci Laboratory Solutions B.V.
	Postbus 12103,
	3004 GC Rotterdam, the Netherlands
Pharmacy < <i>if applicable</i> >	Not applicable

PROTOCOL SIGNATURE SHEET

Name	Signature	Date
Head of Department Gastroenterology & Hepatology:		30/
Prof. M.J. Bruno, MD, PhD	2	6 /
Erasmus MC University Medical Centre, Rotterdam		1020
Principal Investigator:		
Prof. M.J. Bruno, MD, PhD		7.1
Head of Gastroenterology & Hepatology department,		6 /
Erasmus MC University Medical Centre, Rotterdam		1.20

TABLE OF CONTENTS

1.	INT	RODUCTION AND RATIONALE	11
2.	OB	JECTIVES	12
3.	ST	UDY DESIGN	12
4.	ST	UDY POPULATION	14
	a. F	Population (base)	14
	b. I	nclusion criteria	14
	c. E	Exclusion criteria	14
	d. S	Sample size calculation	14
5.	TR	EATMENT OF SUBJECTS	16
	5.1	Investigational product/treatment	16
	5.2	Use of co-intervention (if applicable)	16
	5.3	Escape medication (if applicable)	17
6	IN\	ESTIGATIONAL PRODUCT	18
	6.1	Name and description of investigational product(s)	18
	Not a	pplicable	18
	6.2	Summary of findings from non-clinical studies	18
	6.3	Summary of findings from clinical studies	18
	6.4	Summary of known and potential risks and benefits	18
	6.5	Description and justification of route of administration and dosage	18
	Not a	pplicable	18
	6.6	Dosages, dosage modifications and method of administration	18
	Not a	pplicable	18
	6.7	Preparation and labelling of Investigational Medicinal Product	18
	Not a	pplicable	18
	6.8	Drug accountability	18
	Not a	pplicable	18
7	NO	N-INVESTIGATIONAL PRODUCT	19
	7.1	Name and description of non-investigational product(s)	19
	7.2	Summary of findings from non-clinical studies	19
	7.3	Summary of findings from clinical studies	19
	7.4	Summary of known and potential risks and benefits	19
	7.5	Description and justification of route of administration and dosage	20
	7.6	Dosages, dosage modifications and method of administration	20
	7.7	Preparation and labelling of Non Investigational Medicinal Product	20
	7.8	Drug accountability	20
8	ME	THODS	21
	8.1	Study parameters/endpoints	21
	8.1	.1 Main study parameter/endpoint	21
	8.1	.2 Secondary study parameters/endpoints (if applicable)	21
	Not	t applicable	21
	8.1	.3 Other study parameters (if applicable)	21

	8.2	Randomisation, blinding and treatment allocation	21
	8.3	Study procedures	21
	8.4	Withdrawal of individual subjects	23
	8.4.	1 Specific criteria for withdrawal (if applicable)	24
	Not	applicable	24
	8.5	Replacement of individual subjects after withdrawal	24
	Inclus	ion will end after inclusion of 5 volunteers in each of the six groups	24
	8.6	Follow-up of subjects withdrawn from treatment	24
	Follow	<i>y</i> -up is not performed	24
	8.7	Premature termination of the study	24
9	SAF		25
	9.1	Temporary halt for reasons of subject safety	25
	9.2	AEs, SAEs and SUSARs	25
	9.2.	1 Adverse events (AEs)	25
	9.2.	2 Serious adverse events (SAEs)	25
	9.2.	3 Suspected unexpected serious adverse reactions (SUSARs)	26
	9.3	Annual safety report	26
	9.4	Follow-up of adverse events	
	9.5	Data Safety Monitoring Board (DSMB) / Safety Committee]	
10) S		27
	10.1	Primary study parameter(s)	27
	10.2	Secondary study parameter(s)	27
	10.3	Other study parameters	27
	10.4	Interim analysis (if applicable)	27
11	E	THICAL CONSIDERATIONS	
	11.1	Regulation statement	
	11.2	Recruitment and consent	
	11.3	Objection by minors or incapacitated subjects (if applicable)	
	11.4	Benefits and risks assessment, group relatedness	
	11.5	Compensation for injury	
	11.6	Incentives (if applicable)	
12	2 A	DMINISTRATIVE ASPECTS. MONITORING AND PUBLICATION	
	12.1	Handling and storage of data and documents	
	12.2	Monitoring and Quality Assurance	
	12.3	Amendments	
	12.4	Annual progress report	
	12.5	Temporary halt and (prematurely) end of study report	
	12.6	Public disclosure and publication policy	
13	s s	TRUCTURED RISK ANALYSIS	
	13.1	Potential issues of concern	
	13.2	Synthesis	
14	 R	EFERENCES	

LIST OF ABBREVIATIONS AND RELEVANT DEFINITIONS

ABR	General Assessment and Registration form (ABR form), the application
	form that is required for submission to the accredited Ethics Committee;
	in Dutch: Algemeen Beoordelings- en Registratieformulier (ABR-
	formulier)
AE	Adverse Event
AR	Adverse Reaction
ATP	Adenosine triphosphate
CA	Competent Authority
ССМО	Central Committee on Research Involving Human Subjects; in Dutch:
	Centrale Commissie Mensgebonden Onderzoek
СМІ	Chronic mesenteric ischemia
CV	Curriculum Vitae
DSMB	Data Safety Monitoring Board
EU	European Union
EudraCT	European drug regulatory affairs Clinical Trials
GCP	Good Clinical Practice
GDPR	General Data Protection Regulation; in Dutch: Algemene Verordening
	Gegevensbescherming (AVG)
IB	Investigator's Brochure
IC	Informed Consent
IMP	Investigational Medicinal Product
IMPD	Investigational Medicinal Product Dossier
METC	Medical research ethics committee (MREC); in Dutch: medisch-ethische
	toetsingscommissie (METC)
(S)AE	(Serious) Adverse Event
SPC	Summary of Product Characteristics; in Dutch: officiële
	productinformatie IB1-tekst
Sponsor	The sponsor is the party that commissions the organisation or
	performance of the research, for example a pharmaceutical
	company, academic hospital, scientific organisation or investigator. A
	party that provides funding for a study but does not commission it is not
	regarded as the sponsor, but referred to as a subsidising party.
SUSAR	Suspected Unexpected Serious Adverse Reaction
UAVG	Dutch Act on Implementation of the General Data Protection Regulation;
	in Dutch: Uitvoeringswet AVG

VLS Visible light spectroscopy

WMO Medical Research Involving Human Subjects Act; in Dutch: Wet Medischwetenschappelijk Onderzoek met Mensen

SUMMARY

Rationale: Chronic mesenteric ischemia (CMI) is an incapacitating disease that can progress to potentially fatal acute mesenteric ischemia. The yearly incidence of CMI is 9.2 per 100.000 and will increase due to the aging population and the rising prevalence of cardiovascular risk factors. A gold standard diagnostic test to diagnose CMI is currently lacking, causing both undertreatment and overtreatment of patients and thereby superfluous healthcare expenses. Since oxygen is needed to absorb and metabolise butyrate and glucose in the enterocyte, a ¹³C-butyrate and ¹³C -glucose breath test could theoretically quantify mucosal oxygen content and thereby identify patients with CMI.

Objective: To explore the possibility of detecting mesenteric ischemia by ¹³C-butyrate and ¹³C -glucose breath testing by comparing timing and concentration of the peak of expired ¹³CO₂.

Study design: randomised intervention study

Study population: healthy human volunteers, >18 old

Intervention (if applicable): The intervention group will perform a standardized bicycle ergometry exercise test with a duration of 30 minutes. The aim of the exercise test is to induce temporary mesenteric ischemia.

Main study parameters/endpoints: To explore the possibility of detecting mesenteric ischemia by ¹³C-butyrate and ¹³C -glucose breath testing by comparing timing and concentration of the peak of expired ¹³CO₂.

Nature and extent of the burden and risks associated with participation, benefit and group relatedness: All participating volunteers will perform 2 breath tests at baseline, 0.5, 1, 1.25 1.5, 1.75, 2, 2.25, 2.5, 3, 3.5 and 4 hours after ingestion of ¹³C-butyrate or ¹³C-glucose. A peripheral intravenous cannula will be inserted in patients in the intervention group, in order to obtain a serum blood sample at baseline and after the exercise test. The cannula will also be used to obtain blood for the lactate measurements during the exercise test. Participating volunteers will not experience any benefits of participation. No adverse events are expected and risks associated with participation are deemed very low to non-existent. Insertion of the peripheral intravenous cannula can result in a hematoma.

1. INTRODUCTION AND RATIONALE

Chronic mesenteric ischemia (CMI) is an invalidating disease, causing severe complaints of post-prandial pain, food fear and weight loss(1, 2). When left untreated CMI has a substantial risk of bowel infarction in 26-67% of patients, resulting in high mortality and morbidity (3, 4). Diagnosis of CMI remains difficult, since no gold standard diagnostic test exists. The gold standard diagnosis described in current literature is relief of symptoms after endovascular or surgical revascularisation. In order to take treatment decisions a consensus diagnosis is used. Consensus is reached during a multidisciplinary meeting attended by gastroenterologists, vascular surgeons and interventional radiologists. Consensus is based on history, presence of mesenteric artery stenosis on abdominal imaging and in dedicated centres a functional test, such as visible light spectroscopy (VLS) and tonometry. In patients with a consensus diagnosis of CMI based on a stenosis of a single mesenteric artery, treatment is successful in 73% of patients(5). In the remaining 27% not benefitting from treatment, CMI was never the cause of the patients symptoms.

A more accurate functional test detecting intestinal ischemia is of great value to improve the used consensus diagnosis. Current functional tests have several downsides and are therefore not widely used. VLS, a technique measuring mucosal saturations, is limited by a specificity of 60%, causing a substantial amount of false positive test results(6). Tonometry, a technique measuring luminal carbon dioxide concentrations in stomach and jejunum, has a better specificity of 92%(6). Yet this test will cease to exist in a few years, since tonometry equipment is no longer produced. Stressing the urgency of development of a new and improved functional test.

Breath tests are accurate, simple and widely available tests for diseases, such as helicobacter pylori infection and small intestinal bacterial overgrowth(7, 8). Breath tests using glucose and butyrate could possibly be used to diagnose CMI. Butyrate is metabolized by the intestinal epithelium(9). Metabolism of butyrate is oxygen dependent, since oxidation is one of the steps during the production of adenosine triphosphate (ATP). ¹⁴C-butyrate can be used to measure the oxidation rate of butyrate(10). During the oxidation process the ¹⁴C atom is transferred to an oxygen molecule, producing ¹⁴C-carbon dioxide. The labelled carbon dioxide molecule is exhaled and can be measured in the air exhaled by the patient, thereby quantifying the presence of mucosal oxygen.

Administration of deuterated glucose (¹³C-glucose) is a second potential method to measure mucosal oxygen content. Transport of glucose from the lumen into the epithelial cell is driven by an active process consuming ATP and thus oxygen(11). The transport of glucose is driven by a sodium gradient. At the basolateral side of the cell, Na⁺/K⁺-ATPase extrudes sodium, thereby creating a low sodium concentration within the cell. At the luminal side a Na⁺/glucose cotransporter, driven by the low intracellular sodium concentration, transports sodium and glucose into the cell. Further transport of glucose into the mesenteric circulation occurs by diffusion facilitated by passive glucose transporters. When ¹³C-glucose is metabolized the ¹³C atom is transferred to an oxygen molecule, producing ¹³C-carbon dioxide. The labelled carbon dioxide molecule is exhaled and can be measured in the air exhaled by the patient.

In subjects with low mucosal oxygen concentrations, the peak in exhaled ¹³CO₂ of butyrate and glucose is expected to be delayed and lower compared to control subjects with normal mucosal oxygen concentrations. Thereby creating a functional test measuring mucosal oxygen content, which could be of great value in diagnosing CMI.

2. OBJECTIVES

Primary Objective:

To explore the possibility of detecting mesenteric ischemia by ¹³C-butyrate and ¹³C-glucose breath testing by comparing timing and concentration of the peak of expired ¹³CO₂.

3. STUDY DESIGN

This study is a multi-center randomized interventional proof of principal study, exploring the possibility of quantifying mucosal oxygen content by ¹³C-butyrate and ¹³C-glucose breath testing.

Inclusion

The study is conducted in healthy volunteers without gastrointestinal complaints and unremarkable medical history. Volunteers are recruited by distribution of patient information folders, containing study information and the investigators contact information, at the gastroenterology and Hepatology outpatient clinics of Erasmus MC University Medical Center and Franciscus Gasthuis & Vlietland. When volunteers contact the investigator the study outline is discussed and inclusion and exclusion criteria are checked. Volunteers are asked whether they have a medical history or are currently experiencing any complaints, they do not undergo additional tests or check-ups to ensure they are healthy. When a healthy volunteer decides to participate in the study an Informed Consent Form will be signed.

Control

Two different control groups both consisting of 5 volunteers will be used during the study. The first group will receive ¹³C-butyrate and the second group ¹³C-glucose. The control groups will perform the breath tests without performing any physical exercise.

Intervention

Two different intervention groups both consisting of 5 volunteers will be used during the study. The first group will receive ¹³C-butyrate, the second group ¹³C-glucose. All intervention groups will be performing a standardized bicycle ergometer exercise test with a total duration

of 30 minutes(12). The exercise test has been proven to elicit mesenteric ischemia in athletes. Previous studies suggest an additional 10 minute period of splanchnic hypo perfusion after discontinuation of exercise(13, 14).

The exercise test consist of 3 phases.

- Phase 1: The first 10 minutes of exercise are used to gradually increase the workload until submaximal exercise intensity is reached. Submaximal exercise is defined as lactate between 3 and 5.5 mmol/L. Lactate measurements are performed every 2 minutes, using a rapid lactate measurement kit.
- Phase 2: From minute 10 until 20 submaximal exercise intensity is maintained by adjusting the workload based on lactate measurements. The lactate measurements are performed every 3 minutes.
- Phase 3: Minute 20 until 30 are used to reach maximal exercise intensity. Every 3
 minutes the workload is increased by 10% of the submaximal workload until
 exhaustion. Lactate measurements will be continued with 3 minute intervals.

Randomisation

Allocation to the ¹³C-butyrate or ¹³C-glucose group and allocation to the control or intervention group will be performed by randomisation. Blinding of the healthy volunteers will not be possible, since the intervention group has to perform an exercise test.

Breath test

Both intervention and control group will perform breath tests at the following time points baseline, 0.5, 1, 1.25 1.5, 1.75, 2, 2.25, 2.5, 3, 3.5 and 4 hours after ingestion of ¹³C-butyrate or ¹³C-glucose (figure 1). A breath sample is obtained by blowing a single breath of air through a straw into a test tube. Two breath samples are collected at all mentioned time points. The decision to perform breath testing for a total of 4 hours was based on a study that determined ¹⁴CO₂ elimination patterns after instillation of butyrate in human caecum and a study describing ¹³CO₂ elimination patterns in healthy volunteers(10, 15). The tracer could be measured in exhaled air at the first time point, which was 30 minutes after instillation. A peak in expired ¹⁴CO₂ was measured after 2 hours. Four hours after instillation approximately 55% of the total ¹⁴CO₂ dose had been expired, 24 hours after instillation 70% of the instilled dose had been expired. Since we are interested in the peak of expired ¹³CO₂ a measurement period of 4 hours seems sufficient, even in the intervention group, in whom a delayed or lower peak is expected. The frequency of breath testing is intensified between 1 and 2.5 hours after ingestion, in order to detect the timing of the ¹³CO₂ peak more precisely.

Figure 1. timeline collection of breath samples.



Dosages

All volunteers will be instructed to take their last meal 6 hours before start of the breath test. Volunteers in the ¹³C-butyrate group receive an oral dose of 0.03 µmol ¹³C-butyrate(10). Volunteers in the ¹³C-glucose group receive an oral dose of 20 mmol ¹³C-glucose(16).

Follow-up

There is no follow-up of healthy volunteers.

4. STUDY POPULATION

a. Population (base)

Healthy volunteers with no gastrointestinal complains and unremarkable medical history will be asked to participate in our study through information folders.

b. Inclusion criteria

In order to be eligible to participate in this study, a subject must meet all of the following criteria:

- ≥18 years of age
- Experience with cycling
- Signed informed consent
- Unremarkable medical history (no gastrointestinal diseases, no cardiac or pulmonary diseases)
- No gastrointestinal complaints

c. Exclusion criteria

A potential subject who meets any of the following criteria will be excluded from participation in this study:

- Unable to sign informed consent
- Pregnancy

d. Sample size calculation

This study is a proof of principal study, no previous studies exist on this topic, therefore a sample size calculation could not be performed. We have chosen to include a total of 20 healthy volunteers in order to be able to compare 5 control subjects and 5 intervention

subjects for each breath test. In case the number of 5 subjects per group is insufficient to detect a statistical difference, the generated data can be used to perform a sample size calculation for a second study.

5. TREATMENT OF SUBJECTS

5.1 Investigational product/treatment

Intervention

Two different intervention groups both consisting of 5 volunteers will be used during the study. The first group will receive ¹³C-butyrate and the second group ¹³C-glucose. Both intervention groups will be performing a standardized bicycle ergometer exercise test with a total duration of 30 minutes(12). The exercise test has been proven to elicit mesenteric ischemia in athletes. Previous studies suggest an additional 10 minute period of splanchnic hypo perfusion after discontinuation of exercise(13, 14).

The exercise test consist of 3 phases.

- Phase 1: The first 10 minutes of exercise are used to gradually increase the workload until submaximal exercise intensity is reached. Submaximal exercise is defined as lactate between 3 and 5.5 mmol/L. Lactate measurements are performed every 2 minutes, using a rapid lactate measurement kit.
- Phase 2: From minute 10 until 20 submaximal exercise intensity is maintained by adjusting the workload based on lactate measurements. The lactate measurements are performed every 3 minutes.
- Phase 3: Minute 20 until 30 are used to reach maximal exercise intensity. Every 3 minutes the workload is increased by 10% of the submaximal workload until exhaustion. Lactate measurements will be continued with 3 minute intervals.

Control

Two different control groups both consisting of 5 volunteers will be used during the study. The first group will receive ¹³C-butyrate and the second group ¹³C-glucose. The control groups will perform the breath tests without performing any physical exercise.

Dosages

All volunteers will be instructed to take their last meal 6 hours before start of the breath test. Volunteers in the ¹³C-butyrate group receive an oral dose of 0.03 µmol ¹³C-butyrate(10). Volunteers in the ¹³C-glucose group receive an oral dose of 20 mmol ¹³C-glucose(16).

5.2Use of co-intervention (if applicable)

5.3 Escape medication (if applicable)

6 INVESTIGATIONAL PRODUCT

Not applicable

6.1 Name and description of investigational product(s)

Not applicable

6.2 Summary of findings from non-clinical studies

Not applicable

6.3 Summary of findings from clinical studies

Not applicable

6.4 Summary of known and potential risks and benefits

Not applicable

6.5 Description and justification of route of administration and dosage

Not applicable

6.6 Dosages, dosage modifications and method of administration

Not applicable

6.7 Preparation and labelling of Investigational Medicinal Product

Not applicable

6.8 Drug accountability

7 NON-INVESTIGATIONAL PRODUCT

7.1 Name and description of non-investigational product(s)

¹³C-butyrate is a stable isotope used as a tracer molecule.

https://www.sigmaaldrich.com/catalog/product/aldrich/488380?lang=en®ion=NL

D-Glucose-1-¹³C is a stable isotope used as a tracer molecule.

https://www.sigmaaldrich.com/catalog/product/aldrich/297046?lang=en®ion=NL

7.2 Summary of findings from non-clinical studies

¹³C-butyrate

We refer to the safety information statement on the website of Sigma-Aldrich https://www.sigmaaldrich.com/catalog/product/aldrich/488380?lang=en®ion=NL

D-Glucose-1-13C

We refer to the safety information statement on the website of Sigma-Aldrich https://www.sigmaaldrich.com/catalog/product/aldrich/297046?lang=en®ion=NL

7.3 Summary of findings from clinical studies

¹³C-butyrate

A study including 12 healthy volunteers and 12 Crohn's disease patients showed that ¹³Cbutyrate absorption metabolism can be measured by ¹³CO₂ breath testing(17). They detected ¹³CO₂ at 30 minutes after oral administration of ¹³C-butyrate, with a peak ¹³CO₂ concentration at 1 hour after administration. A study showed similar results and timing of expired ¹⁴CO₂ concentrations after instillation of ¹⁴C-butyrate in the cecum(10).

D-Glucose-1-13C

A study including 8 healthy highly trained volunteers examined the oxidation of 13 C-glucose during exercise(16). This study showed that 13 CO₂ could be measured reliably, in expired air, after administration of a 13 C-glucose dose of 0.05g/kg bodyweight.

7.4 Summary of known and potential risks and benefits

¹³C-butyrate

We refer to the safety information statement on the website of Sigma-Aldrich <u>https://www.sigmaaldrich.com/catalog/product/aldrich/488380?lang=en®ion=NL</u>

D-Glucose-1-13C

We refer to the safety information statement on the website of Sigma-Aldrich <u>https://www.sigmaaldrich.com/catalog/product/aldrich/297046?lang=en®ion=NL</u>

7.5 Description and justification of route of administration and dosage

¹³C-butyrate

A study performing ${}^{14}CO_2$ breath testing after instillation of ${}^{14}C$ -butyrate in the cecum showed that a tracer dose of 0.03 µmol was sufficient and could be detected in expired air(10).

D-Glucose-1-13C

A study performing ¹³CO₂ breath testing during exercise showed that ¹³CO₂ could be measured reliably after administering an oral dose of 0.05g/kg bodyweight ¹³C-glucose(16). This dosage equals approximately 20mmol of glucose, when using a bodyweight of 70kg.

7.6 Dosages, dosage modifications and method of administration

¹³C-butyrate

A study performing ${}^{14}CO_2$ breath testing after instillation of ${}^{14}C$ -butyrate in the cecum showed that a tracer dose of 0.03 µmol was sufficient and could be detected in expired air(10).

D-Glucose-1-13C

A study performing ¹³CO₂ breath testing during exercise showed that ¹³CO₂ could be measured reliably after administering 0.05g/kg bodyweight ¹³C-glucose(16). This dosage equals approximately 20mmol of glucose, when using a bodyweight of 70kg.

Both ¹³C-butyrate and D-Glucose-1-¹³C are administered orally, since metabolism and absorption by the enterocytes are the processes that are of interest.

7.7 Preparation and labelling of Non Investigational Medicinal Product

Preparation and labelling of the non-investigational medicinal products will be done according to the relevant GMP guidelines by licensed staff.

7.8 Drug accountability

¹³C-butyrate and D-Glucose-1-¹³C solutions will be prepared at the Gastroenterology & Hepatology department.

8 METHODS

8.1 Study parameters/endpoints

8.1.1 Main study parameter/endpoint

To explore the possibility of detecting mesenteric ischemia by ¹³C-butyrate and ¹³C-glucose breath testing by comparing timing and concentration of the peak of expired ¹³CO₂.

8.1.2 Secondary study parameters/endpoints (if applicable)

Not applicable

8.1.3 Other study parameters (if applicable)

Not applicable

8.2 Randomisation, blinding and treatment allocation

Participating volunteers will not be blinded. Randomization will be performed into four groups of five volunteers.

Randomisation will be performed through a random list generated by an online randomisation tool and is executed by a person not actively involved in the study. The randomisation tool can be found at <u>https://www.sealedenvelope.com/simple-randomiser/v1/lists</u>. The used seed will be noted and saved, the treatment groups are: BC, BI, GC, GI. Block sizes will be 5 blocks of 4, resulting in a randomized list of 20 subjects (5 per group). The created list will be saved and used to fill 20 envelopes numbered 1-20. Each envelope will contain the treatment group as specified in the randomly generated list. Envelope 1 will be for the first subject enrolled in the study, envelope 2 for the second subject enrolled in the study, etc.

8.3 Study procedures

Inclusion

The study is conducted in healthy volunteers without gastrointestinal complaints and unremarkable medical history. Volunteers are recruited by distribution of patient information folders, containing study information and the investigators contact information, at the gastroenterology and Hepatology outpatient clinics of Erasmus MC University Medical Center and Franciscus Gasthuis & Vlietland. When volunteers contact the investigator the study outline is discussed and inclusion and exclusion criteria are checked. Volunteers are asked whether they have a medical history or are currently experiencing any complaints, they do not undergo additional tests or check-ups to ensure they are healthy. When a healthy volunteer decides to participate in the study an Informed Consent Form will be signed. Volunteers will be randomized and assigned to one of four study groups.

Control

Two different control groups both consisting of 5 volunteers will be used during the study. The first group will receive ¹³C-butyrate and the second group ¹³C-glucose. The control groups will perform the breath tests without performing any physical exercise.

Intervention

Two different intervention groups both consisting of 5 volunteers will be used during the study. The first group will receive ¹³C-butyrate and the second group ¹³C-glucose. All intervention groups will be performing a standardized bicycle ergometer exercise test with a total duration of 30 minutes(12). The exercise test has been proven to elicit mesenteric ischemia in athletes. Previous studies suggest an additional 10 minute period of splanchnic hypo perfusion after discontinuation of exercise(13, 14).

The exercise test consist of 3 phases.

- Phase 1: The first 10 minutes of exercise are used to gradually increase the workload until submaximal exercise intensity is reached. Submaximal exercise is defined as lactate between 3 and 5.5 mmol/L. Lactate measurements are performed every 2 minutes, using a rapid lactate measurement kit.
- Phase 2: From minute 10 until 20 submaximal exercise intensity is maintained by adjusting the workload based on lactate measurements. The lactate measurements are performed every 3 minutes.
- Phase 3: Minute 20 until 30 are used to reach maximal exercise intensity. Every 3 minutes the workload is increased by 10% of the submaximal workload until exhaustion. Lactate measurements will be continued with 3 minute intervals.

All volunteers will be instructed to take their last meal 6 hours before start of the breath test. Volunteers in the ¹³C-butyrate group receive an oral dose of 0.03 µmol ¹³C-butyrate(10). Volunteers in the ¹³C-glucose group receive an oral dose 20 mmol ¹³C-glucose.

Randomisation

Allocation to the ¹³C-butyrate and ¹³C-glucose group and allocation to the control or intervention group will be performed by randomisation. Blinding of the healthy volunteers will not be possible, since the intervention group has to perform an exercise test.

Breath test

Both intervention and control group will perform breath tests at the following time points baseline, 0.5, 1, 1.25 1.5, 1.75, 2, 2.25, 2.5, 3, 3.5 and 4 hours after ingestion of ¹³C-butyrate

or ¹³C-glucose. Two breath samples are collected at all mentioned time points. The decision to perform breath testing for a total of 4 hours was based on a study that determined ¹⁴CO₂ elimination patterns after instillation of butyrate in human caecum and a study describing ¹³CO₂ elimination patterns in healthy volunteers(10, 15). The tracer could be measured in exhaled air at the first time point, which was 30 minutes after instillation. A peak in expired ¹⁴CO₂ was measured after 2 hours. Four hours after instillation approximately 55% of the total ¹⁴CO₂ dose had been expired, 24 hours after instillation 70% of the instilled dose had been expired. Since we are interested in the peak of expired ¹³CO₂ a measurement period of 4 hours seems sufficient, even in the intervention group, in whom a delayed or lower peak is expected. The frequency of breath testing is intensified between 1 and 2.5 hours after ingestion, in order to detect the timing of the ¹³CO₂ peak more precisely.

Follow-up

There is no follow-up of healthy volunteers.

Study procedures	Control	Intervention
	group	group
Oral dose of either 0.03 µmol ¹³ C-butyrate or 20 mmol	Yes	Yes
¹³ C-glucose		
Insertion of venous canule	No	Yes
(measurement of lactate, 2 serum samples: 1 at baseline		
and 1 after the exercise test)		
Bicycle ergometer exercise test	No	Yes
Collection of 2 breath samples at baseline, 0.5, 1, 1.25	Yes	Yes
1.5, 1.75, 2, 2.25, 2.5, 3, 3.5 and 4 hours after ingestion		
of ¹³ C-butyrate or ¹³ C-glucose		

Analysis of breath samples

Measurement of ¹³CO₂ will be performed using mass spectrometry (IDMicro Breath, Compact Science Systems, Newcastle-Under-Lyme, United Kingdom). The mass spectrometry procedure is equal to the procedures used to measure ¹³CO₂ in testing for Helicobacter Pylori, which is current clinical practice.

8.4 Withdrawal of individual subjects

Subjects can leave the study at any time for any reason if they wish to do so without any consequences. The investigator can decide to withdraw a subject from the study for urgent medical reasons.

8.4.1 Specific criteria for withdrawal (if applicable)

Not applicable

8.5 Replacement of individual subjects after withdrawal

Inclusion will end after inclusion of 5 volunteers in each of the four groups.

8.6 Follow-up of subjects withdrawn from treatment

Follow-up is not performed.

8.7 Premature termination of the study

The study will be terminated prematurely when participation is associated with serious health risks, which are not expected.

9 SAFETY REPORTING

9.1 Temporary halt for reasons of subject safety

In accordance to section 10, subsection 4, of the WMO, the sponsor will suspend the study if there is sufficient ground that continuation of the study will jeopardise subject health or safety. The sponsor will notify the accredited METC without undue delay of a temporary halt including the reason for such an action. The study will be suspended pending a further positive decision by the accredited METC. The investigator will take care that all subjects are kept informed.

9.2 AEs, SAEs and SUSARs

9.2.1 Adverse events (AEs)

Adverse events are defined as an undesirable experience occurring to a subject during the study, related to the study. All adverse events reported spontaneously by the subject or observed by the investigator or his staff will be recorded.

9.2.2 Serious adverse events (SAEs)

A serious adverse event is any untoward medical occurrence or effect that

- results in death;
- is life threatening (at the time of the event);
- requires hospitalisation or prolongation of existing inpatients' hospitalisation;
- results in persistent or significant disability or incapacity;
- is a congenital anomaly or birth defect; or
- any other important medical event that did not result in any of the outcomes listed above due to medical or surgical intervention but could have been based upon appropriate judgement by the investigator.

An elective hospital admission will not be considered as a serious adverse event.

The sponsor will report the SAEs through the web portal *ToetsingOnline* to the accredited METC that approved the protocol, within 7 days of first knowledge for SAEs that result in death or are life threatening followed by a period of maximum of 8 days to complete the initial preliminary report. All other SAEs will be reported within a period of maximum 15 days after the sponsor has first knowledge of the serious adverse events.

9.2.3 Suspected unexpected serious adverse reactions (SU-SARs)

Not applicable.

9.3 Annual safety report

Not applicable

9.4 Follow-up of adverse events

All AEs will be followed until they have abated, or until a stable situation has been reached. Depending on the event, follow up may require additional tests or medical procedures as indicated, and/or referral to the general physician or a medical specialist. SAEs need to be reported till end of study within the Netherlands, as defined in the protocol

9.5 [Data Safety Monitoring Board (DSMB) / Safety Committee]

10 STATISTICAL ANALYSIS

Baseline characteristics of the intervention and control group will be shown and compared. Categorical data is compared by chi-square or Fisher exact testing. Numerical data is shown as mean and standard deviation or median and interquartile range. Comparison of the numerical data of intervention and control group will be performed by the independent sample t-test or the Mann-Whitney-U test.

10.1 Primary study parameter(s)

The height of the ¹³CO₂ peak of the intervention and control groups will be compared by the independent sample t-test or Mann-Whitney-U test. Repeated measurement correction will be performed with the Bonferroni method. The area under the ¹³CO₂ curves will be compared by the independent sample t-test or Mann-Whitney-U test.

10.2 Secondary study parameter(s)

Not applicable

10.3 Other study parameters Not applicable

10.4 Interim analysis (if applicable)

11 ETHICAL CONSIDERATIONS

11.1 Regulation statement

The study will be performed according to the principles of the Declaration of Helsinki, of Good Clinical Practice, and the applicable laws and regulations of the Netherlands, including but not limited to the Medical Research Involving Human Subjects Act (WMO) and the Personal Data Protection Act.

11.2 Recruitment and consent

Volunteers are recruited by distribution of patient information folders, containing study information and the investigators contact information, at the gastroenterology and Hepatology outpatient clinics of Erasmus MC University Medical Center and Franciscus Gasthuis & Vlietland. When volunteers contact the investigator the study outline is discussed and inclusion and exclusion criteria are checked. Volunteers are asked whether they have a medical history or are currently experiencing any complaints, they do not undergo additional tests or check-ups to ensure they are healthy. When a healthy volunteer decides to participate in the study an Informed Consent Form will be signed.

11.3 Objection by minors or incapacitated subjects (if applicable)

Not applicable

11.4 Benefits and risks assessment, group relatedness

Participation in this study will not be beneficial for the healthy volunteers. However, this study will provide a valuable insight into the feasibility of ¹³C-butyrate and ¹³C-glucose breath testing. This proof-of-principal study is the first step in the development of a more accurate and less cumbersome and less invasive functional test for the detection of CMI.

There are no known side-effects of the administration of ¹³C-glucose. Administration of high concentrations of ¹³C-butyrate could result in diarrhoea, however tracer dosages are used in this study.

11.5 Compensation for injury

The sponsor/investigator has a liability insurance which is in accordance with article 7 of the WMO.

The sponsor (also) has an insurance which is in accordance with the legal requirements in the Netherlands (Article 7 WMO). This insurance provides cover for damage to research subjects through injury or death caused by the study.

The insurance applies to the damage that becomes apparent during the study or within 4 years after the end of the study.

11.6 Incentives (if applicable)

12 ADMINISTRATIVE ASPECTS, MONITORING AND PUBLICATION

12.1 Handling and storage of data and documents

The handling of personal data will be compatible with the Code of Good Behaviour (Code goed gedrag) and the Dutch Personal Data Protection Act (Wet Bescherming Persoonsgegevens). The collected data will be recorded in an electronic database, which contains all relevant clinical data of the volunteers. This database will be provided with a password, which is only available for the investigators. The database will be anonymized by replacing all names of volunteers by a code and will be used for statistical analysis. Only these codes will be used for references in reports and publications about this investigation. Volunteers will be asked for their permission to store the material for 15 years and that it could be used for scientific research in the future with the same goal.

12.2 Monitoring and Quality Assurance

Not applicable

12.3 Amendments

Amendments are changes made to the research after a favourable opinion by the accredited METC has been given. All amendments will be notified to the METC that gave a favourable opinion.

12.4 Annual progress report

The sponsor/investigator will submit a summary of the progress of the trial to the accredited METC once a year. Information will be provided on the date of inclusion of the first subject, numbers of subjects included and numbers of subjects that have completed the trial, serious adverse events/ serious adverse reactions, other problems, and amendments.

12.5 Temporary halt and (prematurely) end of study report

The investigator/sponsor will notify the accredited METC of the end of the study within a period of 8 weeks. The end of the study is defined as the last patient's last visit.

The sponsor will notify the METC immediately of a temporary halt of the study, including the reason of such an action.

In case the study is ended prematurely, the sponsor will notify the accredited METC within 15 days, including the reasons for the premature termination.

Within one year after the end of the study, the investigator/sponsor will submit a final study report with the results of the study, including any publications/abstracts of the study, to the accredited METC.

12.6 Public disclosure and publication policy

Results of the study will be published in an international journal. There are no limitations or restrictions for publication mention the arrangements made between the sponsor and the investigator concerning the public disclosure and publication.

STRUCTURED RISK ANALYSIS

12.7 Potential issues of concern

<u>a. Level of knowledge about mechanism of action</u> Both ¹³C-butyrate and ¹³C-glucose are stable isotopes

b. Previous exposure of human beings with the test product(s) and/or products with a similar biological mechanism

¹³C-butyrate and ¹³C-glucose have been administered in human subjects(10). Adverse events were not reported.

<u>c. Can the primary or secondary mechanism be induced in animals and/or in *ex-vivo* human cell material? Not applicable</u>

<u>d. Selectivity of the mechanism to target tissue in animals and/or human beings</u> Both ¹³C-butyrate and ¹³C-glucose can be metabolized in cells other than enterocytes, yet in this case the timing of the peak is expected to be delayed, since absorption and transportation through the bloodstream are required.

e. Analysis of potential effect

A study performing ¹⁴CO₂ breath testing after instillation of ¹⁴C-butyrate in the cecum showed that a tracer dose of 0.03 µmol was sufficient and could be detected in expired air(10). A study has shown that a ¹³C-glucose dose of 0.05 gr/kg bodyweight (approximately 3.5gr in an individual of 70 kg) could be detected in expired air. The effects of exercise induced mesenteric ischemia on the breath test patterns have not been studied.

f. Pharmacokinetic considerations

Not applicable

g. Study population

Heathy volunteers have been used in previous studies administering ¹³C-butyrate and ¹³Cglucose.

h. Interaction with other products

No drug interactions are expected, since ¹³C-butyrate and ¹³C-glucose are metabolized by the enterocytes.

i. Predictability of effect

A study that determined ${}^{14}CO_2$ elimination patterns after instillation of butyrate in human caecum and a study described ${}^{13}CO_2$ elimination patterns in healthy volunteers(10, 15). The tracer could be measured in exhaled air at the first time point, which was 30 minutes after instillation. A peak in expired ${}^{14}CO_2$ was measured after 2 hours. Four hours after instillation approximately 55% of the total ${}^{14}CO_2$ dose had been expired, 24 hours after instillation 70% of the instilled dose had been expired. Since we are interested in the peak of expired ${}^{13}CO_2$ a measurement period of 4 hours seems sufficient, even in the intervention group, in whom a delayed or lower peak is expected.

j. Can effects be managed?

No effects are expected, but when unexpected effects do occur the exercise test can be stopped.

12.8 Synthesis

Not applicable

13 REFERENCES

1. Alahdab F, Arwani R, Pasha AK, Razouki ZA, Prokop LJ, Huber TS, et al. A systematic review and meta-analysis of endovascular versus open surgical revascularization for chronic mesenteric ischemia. J Vasc Surg. 2018.

2. Mensink PB, Moons LM, Kuipers EJ. Chronic gastrointestinal ischaemia: shifting paradigms. Gut. 2011;60(5):722-37.

3. Thomas JH, Blake K, Pierce GE, Hermreck AS, Seigel E. The clinical course of asymptomatic mesenteric arterial stenosis. J Vasc Surg. 1998;27(5):840-4.

4. Kolkman JJ, Mensink PB, van Petersen AS, Huisman AB, Geelkerken RH. Clinical approach to chronic gastrointestinal ischaemia: from 'intestinal angina' to the spectrum of chronic splanchnic disease. Scand J Gastroenterol Suppl. 2004(241):9-16.

5. van Dijk LJD, Moons LMG, van Noord D, Moelker A, Verhagen HJM, Bruno MJ, et al. Persistent symptom relief after revascularization in patients with single-artery chronic mesenteric ischemia. J Vasc Surg. 2018.

6. van Noord D, Kolkman JJ. Functional testing in the diagnosis of chronic mesenteric ischemia. Best Pract Res Clin Gastroenterol. 2017;31(1):59-68.

7. Rezaie A, Buresi M, Lembo A, Lin H, McCallum R, Rao S, et al. Hydrogen and Methane-Based Breath Testing in Gastrointestinal Disorders: The North American Consensus. Am J Gastroenterol. 2017;112(5):775-84.

8. Malfertheiner P, Megraud F, O'Morain CA, Gisbert JP, Kuipers EJ, Axon AT, et al. Management of Helicobacter pylori infection-the Maastricht V/Florence Consensus Report. Gut. 2017;66(1):6-30.

9. Bergman EN. Energy contributions of volatile fatty acids from the gastrointestinal tract in various species. Physiol Rev. 1990;70(2):567-90.

10. Hoverstad T, Bohmer T, Fausa O. Absorption of short-chain fatty acids from the human colon measured by the 14CO2 breath test. Scand J Gastroenterol. 1982;17(3):373-8.

11. Chen L, Tuo B, Dong H. Regulation of Intestinal Glucose Absorption by Ion Channels and Transporters. Nutrients. 2016;8(1).

12. ter Steege RW, Geelkerken RH, Huisman AB, Kolkman JJ. Abdominal symptoms during physical exercise and the role of gastrointestinal ischaemia: a study in 12 symptomatic athletes. Br J Sports Med. 2012;46(13):931-5.

13. Otte JA, Oostveen E, Geelkerken RH, Groeneveld AB, Kolkman JJ. Exercise induces gastric ischemia in healthy volunteers: a tonometry study. J Appl Physiol (1985). 2001;91(2):866-71.

14. Qamar MI, Read AE. Effects of exercise on mesenteric blood flow in man. Gut. 1987;28(5):583-7.

15. Tanaka K, Matsuura T, Shindo D, Aida Y, Matsumoto Y, Nagatsuma K, et al. Noninvasive assessment of insulin resistance in the liver using the fasting (13)C-glucose breath test. Transl Res. 2013;162(3):191-200.

16. Leijssen DP, Saris WH, Jeukendrup AE, Wagenmakers AJ. Oxidation of exogenous [13C]galactose and [13C]glucose during exercise. J Appl Physiol (1985). 1995;79(3):720-5.

17. Roda A, Simoni P, Magliulo M, Nanni P, Baraldini M, Roda G, et al. A new oral formulation for the release of sodium butyrate in the ileo-cecal region and colon. World J Gastroenterol. 2007;13(7):1079-84.